# Roosevelt Biosafety Training

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## Objectives

- Identify risks and hazards in biological laboratories
- Understand biosafety levels for laboratories and the proper procedures for working in them
- \* Learn ways to reduce risks and hazards by understanding the use of aseptic technique and proper use of biological safety cabinets
- Learn proper procedures for cleaning biohazardous spills
- \* Learn proper disposal procedures for biohazardous waste in biological laboratories

#### \* Risk assessment:

- \* What are the biological and physical hazards of the organism/agent?
- \* What procedures may spread the organism/agent?
- \* What is the best method for inactivating and containing the organism/agent?
- \* What is the pathogenicity of the organism/agent?
- \* What are the potential deficiencies in practices of lab workers?

- \* Risk assessment:
  - Identify agent hazards
    - \* CDC and WHO have guidelines
    - \* Biosafety level
  - \* Identify laboratory procedure hazards
    - \* Aerosols
    - Volume and concentration
    - \* Use of sharps or animals
    - \* Complexity of experiment
  - Determine appropriate biosafety level and additional precautions
  - Evaluate staff and equipment for safe practices
  - \* Review risk assessment with biosafety professionals

#### \* Risk Groups

- \* Organisms are categorized by:
  - Potential effect of the agent on a healthy human adult\*
  - \* Ease and route of transmission
  - \* Infective dose
  - \* Stability in environment
- Knowing the risk group of the organism can determine the procedures necessary for reducing risk
- \* May differ between strains of the same organism, depending on pathogenicity of the different strains
- NIH and WHO definitions differ slightly
- Do not equal biosafety levels (though they are correlated)

## Risk Groups

- Risk Group 1
  - \* Agents not associated with disease in humans or animals
- \* Risk Group 2
  - \* Agents associated with human or animal disease, but are unlikely to cause serious hazard to lab workers
  - \* If exposure happens, preventive or therapeutic interventions are usually available
- \* Risk Group 3
  - \* Agents associated with serious or lethal human or animal disease
  - \* Treatments may exist, and the agent isn't easily spread
  - Use is discouraged at Roosevelt University
- Risk Group 4
  - Agents associated with serious or lethal human or animal disease that can be easily passed between individuals
  - Treatments are not usually available
  - \* Use is prohibited at Roosevelt University

- \* Routes of laboratory transmission
  - Inoculation from contaminated sharps
  - \* Spills and splashes onto skin and mucous membranes
  - Ingestion (mouth pipetting)
  - Animal bites and scratches
  - \* Inhalation of infectious aerosols
    - \* Considered to be a serious hazard and likely culprit of most laboratory exposures

#### Reducing Risks: Aerosols

- \* Procedures that produce aerosols:
  - \* Pipetting
  - \* Blending
  - Centrifugation
  - \* Sonicators and vortex mixers
- \* Vigilant workers can reduce the amount of aerosols by being cautious with above procedures
- \* Use biological safety cabinet when these procedures must be performed

# Reducing Risks: PPE

- \* Minimum Personal Protective Equipment:
  - Safety glasses
  - \* Gloves
  - \* Lab coat
- \* Other equipment that may be necessary based on risk assessment:
  - Mask or face shield
  - \* Respirator
  - \* Gown

# Reducing Risk: Equipment

- Biological Safety Cabinets (BSC)
  - \* Produce a sterile field to protect both the worker and the culture
- \* Centrifuge safety cups
- \* Sealed rotors
- \* Maintenance of equipment is essential

# Reducing Risk: Facility

- Engineering controls to prevent release of hazards
  - \* Directional airflow in the lab
  - Substitution of lower risk agent for higher risk agent if possible
- \* Limited access to building or labs
- \* Training

## Biosafety Level 1

- Well characterized strains of agents not known to cause disease in healthy adults
- Basic containment with sink for hand washing
- \* Laboratory doors kept closed during experiments
- \* Decontaminate work surfaces daily and after spills
- \* All waste decontaminated before disposal
- Mouth pipetting prohibited
- \* Eating, drinking, smoking, and applying cosmetics are not permitted in lab
- \* Wash hands after handling materials and when leaving the lab
- PPE use recommended (required at Roosevelt)
- \* Spills reported to lab manager to ensure proper documentation and clean-up

## Biosafety Level 2

- \* All as in BSL-1
- Moderate-risk agents
- \* Can be used on the open bench if risk of aerosols and splashes is low, higher risk organisms must be manipulated in BSC
- \* PPE required, cannot be worn in non-laboratory areas
- Access to sink for hand washing and decontamination
- \* Access to laboratory is limited or restricted
  - \* Persons with increased risk of acquiring infection or for whom infection is unusually hazardous should not be allowed into labs
  - \* Only individuals that meet entry requirements (e.g. immunizations) may enter
- Workers advised of potential hazards and given proper training in handling agents
- \* Biohazard signs clearly posted on doors and equipment
- \* Sharps use minimized

# Biosafety Level 3

- \* All as in BSL-1 and -2
- \* Agents with a potential for respiratory transmission with serious or lethal infections
- \* Work is discouraged at Roosevelt
- \* Lab personnel must have specific training in handling agents and are supervised while conducting experiments
- \* All work done in BSC or other enclosed equipment
- \* Controlled access to lab and special ventilation to prevent accidental release
- \* Lab must have specific design and containment equipment
  - \* Air lock, shower, or changing room required between unrestricted areas and lab
  - \* Surfaces of walls, floors, and ceilings must be water resistant
  - \* Windows are closed and sealed
  - Exhaust system provided to prevent release of agents
- \* PPE required and may include respirator, double gloves, gowns, etc. Lab coats are not suitable
- \* Workers must comply with entry and exit procedures
- Vacuum lines protected with HEPA filters and liquid traps

## Biosafety Levels 4

- \* All as in BSL-1, -2, and -3
- \* Work is prohibited at Roosevelt
- \* Agents with high risk of lethal disease, easily transmitted (aerosol), with little or no treatment options
- \* Class III BSC or full-body, air-supplied positive pressure suit required for working
- \* Controlled access to labs, specialized ventilation, and waste management systems

#### **Biosafety Cabinets**

- \* Use when procedures are likely to produce aerosols or when high concentrations or large volumes of agent are being used
- Different models provide protection in different ways; must be chosen according to needs of the lab and agents used
- \* Lab workers must be trained in proper use of BSC

#### **Biosafety Cabinets**

\* For proper use of BSC, watch following video:

http://www.youtube.com/watch?v=ZnUW1N-JJz8

- \* Lab workers may require in-person training depending on procedures, experience, and agents being used
- \* Instructors MUST train their students (students cannot train other students)
- In-person training will be provided by instructor or laboratory manager

# Aseptic Technique

- \* Method of laboratory work that prevents contamination by (unwanted) microorganisms
- \* Provides barrier between sterile cell cultures and microorganisms in the environment
- \* Varies depending on whether working on the bench or in a BSC

# Aseptic Technique

\* For proper bench-top aseptic technique, watch the following video:

http://www.youtube.com/watch?v=bRadiLXkqoU

- \* Lab workers may require in-person training depending on procedures, experience, and agents being used
- \* Instructors MUST train their students (students cannot train other students)
- In-person training will be provided by instructor or laboratory manager

- \* Basic biological spill kit should contain:
  - \* Disinfectant (e.g. bleach 1:10 dilution, diluted quaternary solution, or other suitable disinfectant)
  - \* Absorbent material (paper towels, spill pillows, etc.)
  - \* Waste container (biohazard bags and sharps containers)
  - \* PPE
  - \* Mechanical tools (forceps, dustpan and broom)
- \* Procedure can depend on agent spilled and where

- \* BSL-1 spills:
  - Notify others
  - \* Wear PPE
  - \* Surround spill with disinfectant
  - Clean up with paper towels (if large, use spill pillows)
  - \* Re-apply disinfectant to the surface and let sit for 10 minutes. Clean again.
  - Put contaminated waste in biohazard bags for autoclaving
  - \* Wash hands
  - Notify lab manager to assure proper cleanup

#### \* BSL-2 spills:

- \* Evacuate the room and close doors; notify lab manager
- \* Remove any contaminated clothing and decontaminate body surfaces
- Allow at least 30 minutes for potential aerosols to be reduced before reentering
- Don protective clothing and respiratory protective equipment
- \* Decontaminate spill with appropriate disinfectant and allow 10 minutes of contact time
- Clean spill with paper towels or spill pillows and dispose in biohazard bag
  - \* Pick up sharps with forceps or tweezers, never with hands, and dispose of in autoclavable sharps container
- Reapply disinfectant and clean after 10 minutes.
- Wash hands and/or shower after cleaning spill

- \* Biosafety Cabinet Spill
  - Keep cabinet running during the cleanup
  - Remove any contaminated PPE and replace with clean
  - \* Apply appropriate disinfectant to the spill (bleach can be used but should be used with caution; it will corrode the stainless steel)
  - \* Wipe up spill and dispose of paper towels in biohazard bag
  - \* Reapply disinfectant and clean again
  - \* If bleach is used, clean the surface of the cabinet with water to remove traces of bleach
  - Items that must be removed should also be decontaminated before unloading from cabinet
  - Run UV/germicidal lamp for at least 15 minutes for final decontamination (formaldehyde gas can also be used)

#### Waste Disposal

#### \* Sharps

- \* Items capable of puncturing, cutting, or abrading the skin (e.g. broken glass or plastic ware, scalpels, razor blades, needles, etc.)
- \* Never place sharps in regular trash
- \* Dispose of in puncture proof containers
- \* Clean broken glass can go into broken glass containers
- Any sharp contaminated with blood or other biohazard must be decontaminated (autoclave or bleach) and disposed of in an appropriate container
  - \* Leak proof, rigid, puncture-resistant
  - Tightly sealed
  - Labeled with biohazard symbol

#### Waste Disposal

#### \* Biohazardous waste

- \* Waste containing infectious or potentially infectious substances (e.g. blood, bacterial cultures, liquid waste from cell culture, etc.)
- \* All waste must be disposed of in bags marked with biohazard symbol; bags can go into labeled, leak-proof containers to await autoclaving
- Autoclaved before disposal in regular trash
- \* Autoclave should be checked regularly for proper functioning (reaches temperature and pressure, etc.)

# Obtaining Biohazardous Materials

- \* Lab Manager approval required for new organisms
- \* Check risk group and recommended biosafety level
- \* Determine if necessary or if a lower-risk organism can be used instead
- Submit for lab manager approval
  - Will check requirements to determine if Roosevelt has appropriate facilities
  - Currently no facilities for BSL-3
  - Fill out required paperwork in order to obtain organism
- Training with new organism must be conducted by lab manager or instructor

#### Resources and Sources

- \* Center for Disease Control's Biosafety site www.cdc.gov/biosafety
- \* NIH Office of Science Policy for Biosafety http://osp.od.nih.gov/office-biotechnologyactivities/biosafety/nih-guidelines
- \* Roosevelt's CHP https://www.roosevelt.edu/~/media/Files/pdfs/Policies/ /Safety/ChemicalHygienePlan.ashx